

OFFLU Swine Influenza Virus Regional Meeting 20<sup>th</sup> October 2018 XXIX Brazilian Virology Congress and XIII Mercosul Virology Meeting Gramado, RS, Brazil

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### Guatemala



**CENTRO DE** E**STUDIOS EN SALUD** INSTITUTO DE INVESTIGACIONES



### Introduction

- Guatemala is the largest swine producer in Central America (~1.5 millions annually)
- 66% of production in commercial farms
- 34% backyard production
- 1.7% GDP 15.8% agricultural gross domestic product



### Swine IAV in Guatemala

Year	Type of study	Results
2010	<ul> <li>USAC Vet school</li> <li>Swine IAV serological survey among commercial and backyard pig populations</li> <li>Whole country (2155 samples)</li> <li>HI assay</li> </ul>	<ul> <li>H1N1: 20.6% commercial, 41% backyard</li> <li>H3N2: 6.59% commercial, 17% backyard</li> </ul>
2010-2011	<ul><li>CES-UVG</li><li>Two nationwide swine IAV surveys</li><li>Virological and serological assays</li></ul>	<ul> <li>Swine IAV: 15.7% of sampled pigs (30.6% of herds) in 2010 and 11.7% (24.2% of herds) in 2011</li> <li>Seropositivity: 10.6% (16.1% of herds) in 2010 and 1.4% (3.1% of herds) in 2011</li> </ul>
2012, 2014	<ul> <li>CES-UVG</li> <li>Cross-sectional surveys in pigs from peridomestic smallholdings in 2 sites</li> <li>200 samples/site/year</li> <li>Virological and serological assays</li> </ul>	<ul> <li>Swine IAV: 16.5% (2012) and 12% (2014) of sampled pigs</li> <li>Seropositivity: 4.5% (2012) and 1% (2014) of sampled pigs</li> </ul>
2016-2018	<ul> <li>CES-UVG</li> <li>Two-years active swine IAV surveillance in a commercial farm (southern region of Guatemala)</li> <li>2,094 nasal swabs (weekly collection)</li> <li>Virological assays</li> </ul>	<ul> <li>Estimated prevalence among pigs with respiratory signs was 11.1%</li> <li>Only pandemic H1N1 was identified by RRT-PCR or sequencing</li> </ul>





# Isolation and genetic characterization in Guatemala

- 2010-2011: Three pandemic H1N1 and one seasonal human-like H3N2 virus were isolated and sequenced.
  - All gene segment of the H1N1 viruses shared >98% sequence identity with the pandemic lineage.
  - The H3N2 was closely related to human viruses that circulated in Central America in 2010 (distinct to human seasonal vaccine lineage).
- 2016-2018: full genome amplification was performed directly from swabs in 140 RRT-PCR IAV-positive swabs and sequenced by NGS.
  - Only pandemic H1N1 subtype was identify. Data analysis is ongoing.
  - Other additional 141 RRT-PCR IAV-positive swabs samples are being amplified in order to submit them for NGS sequencing.





### Main results and future projects

- Suveillance since 2010 in swine populations
  - Virological detection of IAV
    - 15.7% (2010), 11.7% (2011), 12% (2014) and 13.4% (2016-2018) of sampled pigs
    - Evidence of circulation of IAV of human origin in pigs
    - Phylogenetic analysis of sequences is pending for last period
  - Serological detection of IAV
    - 10.6% (2010), 1.4% (2011) and 1% (2014)
    - Antibodies against viruses from different genetic cluster were detected
- Commercial farms, animal health status and age are potential risk factors associated with IAV infection and exposure.
- Future projects include
  - 3<sup>rd</sup> nation-wide cross-sectional survey at commercial farms level to update information of circulating subtypes
  - Increase number of isolates for better understanding of the evolution and epidemiology of AIV
  - Contribute to establish a network of sentinel surveillance sites and its link with human disease





### Contribution of the team to the group

- Consolidate surveillance in pigs in Guatemala and support other countries in the region.
- Share isolates for regional studies.
- Network with animal and human health authorities.
- Provide training to regional stakeholders.





- Need to develop a sustainable model for IAV surveillance in swine populations with the participation of multiple stakeholders.
- Surveillance data needs to inform producers and health authorities for the control of influenza to prevent and mitigate animal and human diseases.





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#### Department of Population Health, University of Georgia

- Daniel R. Perez
- Lucia Ortiz
- Lab members







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### Who are we?

- Rajao/Perez lab at UGA
  - Daniel R. Perez, Professor
  - Daniela Rajao, Assistant Professor
  - 3 undergraduate students, 4 graduate students, 4 post-docs, 1 lab manager/NGS coordinator
  - Lab is member of the Center for Research on Influenza Pathogenesis (CEIRS-NIAID-NIH)





#### **Center for Research on Influenza Pathogenesis**

- Adolfo Garcia-Sastre, PI, ISMMS NY, P. Palese, N. Bouvier, F. Krammer, R. Albrecht, A Fernandez-Sesma, M. Shaw, H. van Bakel
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- R. Medina, U. Catolica, Chile
- G. Real-Soldevilla, INIA, Spain







### What do we do?

- Support surveillance activities in Argentina and Guatemala through FFS agreements
- Surveillance activities support avian and swine influenza surveillance
- Provide virus characterization support and NGS sequencing capacity to collaborators in Argentina and Guatemala (but not limited to only these two countries)
- Antigenic characterization and Vaccine development (emphasis on swine and avian influenza)





### DIGS – NGS sequencing capacity

A summary of all NGS activity in the Rajao/Perez lab sequencing core since its initiation reflects the an annual sequencing capacity of almost 500 samples.



Validation

Other Lab Genomes Surveillance Genomes

DIGS: Distributed Influenza Genome Sequencing



## Substitutions in key positions on the HA recapitulate antigenic properties (HI profile) of swine H3 influenza viruses

**Collaboration with Amy Vincent and Nicola Lewis** 



Abente, Santos et al, 2016

A/turkey/Ohio/303153/2004 (H3N2) OH/04 – swine-origin TRIG virus





## VACCINE PLATFORMS FOR

	Ö	subunit	Ö
	WIV	VECTORED	LAIV
Delivery route	IM	IM/IN	IN
HI response	+++	++	+
Antibody secreting cells	++	+	+
Memory B cells	+	+	+
Nasal IgA	-/+	-/+	+++
NA antibody	+++	-/+	++
CD4 T cells	++	++	+++
CD8 T cells	-	+	+
Cross protective immunity	-/+	+	++
VAERD	yes	-/+	no

LAIVs have the potential to provide a multidimensional response (humoral – IgA and IgG - and cellular) unparalleled by any other vaccine approach.



From Vincent, Perez, et al 2017, Vet. Micro.





LAIV vs Adjuvanted-WIV vaccine against challenge with contemporary, antigenically drifted H3N2 IAV-S – Study design

Collaboration with Eugenio Abente and Amy Vincent at NADC, USDA-ARS and Daniela Rajao at UGA

	Groups	Vaccine WIV/LAIV (antigenic cluster)	Challenge virus (antigenic cluster)	Number of pigs per group
STUDY 1	NV-NC	No vaccine	No challenge	8
	NV-IN/13	No vaccine	IN/13 (red)	8
	OH/04WIV-IN/13	OH/04WIV (cyan)	IN/13 (red)	8
	OH/04LAIV-IN/13	OH/04LAIV (cyan)	IN/13 (red)	8
STUDY 2	NV-NC	No vaccine	No challenge	5
	NV-IA/14	No vaccine	IA/14 (green)	10
	IA/14WIV-IA/14	IA/14WIV (green)	IA/14 (green)	10
	IA/14LAIV-IA/14	IA/14LAIV (green)	IA/14 (green)	10
	NV-NY/11	No vaccine	NY/11 (red)	9
	IA/14WIV-NY/11	IA/14WIV (green)	NY/11 (red)	10
	IA/14LAIV-NY/11	IA/14LAIV (green)	NY/11 (red)	10

NV, non-vaccinated. NC, non-challenged.

If you like to know the outcome of these studies, please check the paper by Abente, Rajao *et al*, 2018, JVI





### **Contribution to the group**

- Biological and antigenic characterization of viruses
  - Currently producing recombinant viruses for N2 antigenic characterization
- NGS sequencing capacity open to anyone
  - Moving on to nanopore technology and protocols to perform sequencing on site and produce results in real time
- Reverse genetics support

Virus is not needed, only sequence





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