Regional lists of viruses and appropriate diagnostic tests

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# •Select Appropriate Pig with fever, serous nasal discharge, +/- cough

- Flu A Matrix RRT-PCR test (Spackman)
- Routine and commercially available
  - Sensitive 98% and Specific 98%
- Sample types:
  - Respiratory tract samples (tissues or secretions)
  - Bronchial swabs
  - Nasal swabs
  - Oral fluids
  - Tracheal swabs
  - Aerosols
  - Water
  - Environment

# •Flu A Matrix Real-Time RT-PCR test

•More sensitive than NP PCR or Immunoassay or Histopathology or IHC or VI

- •Semi-quantitative
  - (Ct values → lower Ct = higher quantity of viral RNA)

#### •Which Sample Type Is Best?



#### ISU-VDL and UMN-VDL Flu Surveillance (2 years of shared data)

		Flu A Matrix PCR				Total	Ct<35
		Ct<25	Ct 25-<30	Ct 30-<35	Ct 35-<40		
Lung	# VI attempted	353	185	121	336	995	659
	# VI Positive	334	158	35	13	540	527
	VI Success rate	94.6%	85.4%	28.9%	3.9%	54.3%	80.0%
Nasal Swab	# VI attempted	156	147	160	8	471	463
	# VI Positive	129	91	37	0	257	257
	VI Success rate	82.7%	61.9%	23.1%	0.0%	54.6%	55.5%
Oral Fluid	# VI attempted	36	62	119	0	217	217
	# VI Positive	15	17	11	NA	43	43
	VI Success rate	41.7%	27.4%	9.2%	NA	19.8%	19.8%

SIV VI success rate especially on oral fluid is relatively low and needs improvement

### •U of MN VDL 2013 numbers



	Lung	Nasal Swab	Oral Fluids
# of Flu PCRs	80	5211	6358
% Positive`	49%	10%	17%
Average Ct Value	23.04	27.59	29.06
% VI Positive	75%	82%	15%



## Detection of Influenza A virus in pigs

- Proper pig selection and submission of respiratory tract samples or secretions best for virus isolation and subsequent full characterization.
- Oral fluids very useful for detection and subtyping and reveals herd status (pos/neg)quickly/easily

### Serology

- Consistency of ELISA data provides a reliable basis for monitoring herd exposure
- Variety of strains currently circulating in swine dictates need for use of HI testing to monitor/confirm field virus presence

### THANK YOU